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Here we report how induction based fluidics (IBF) has been used to increase analysis speed, sensitivity of MS analysis pictorially and in detail by MALDI and ESI for proteins and peptides, oligonucleotides, metals (Lanthanides and Actinides), explosives and drugs of abuse, and other analyses using IBF, as we discuss the attributes of droplet and related analysis.

Figure 1 is a line graph showing the relationship between the effective field of view (in degrees) and the distance (in meters) for a given sensor. The x-axis is labeled "Effective field of view (m/degree)" and ranges from 0 to 10000. The y-axis is labeled "Distance (m)" and ranges from 0 to 40000. A blue line with circular markers shows a linear relationship, starting at (0,0) and ending at approximately (10000, 40000).

Induction Based Radiolysis (IBR) for Mass Spectrometric Analysis of Organochlorines

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Overview

Mass spectrometry (MS) is a powerful tool for the analysis of organochlorine compounds. However, the analysis of these compounds by MS is often complicated by the presence of numerous isomers and by the presence of numerous interfering ions.

Introduction

Organochlorine compounds are a class of chemicals that have been widely used in industry and agriculture. They are known to be persistent in the environment and to have a variety of adverse effects on human health and the environment. The analysis of these compounds by MS is often complicated by the presence of numerous isomers and by the presence of numerous interfering ions.

Materials and Methods

Organochlorine compounds were analyzed using a gas chromatograph-mass spectrometer (GC-MS). The GC-MS was equipped with a DB-5MS column (30 m x 0.25 mm i.d., 0.1 µm film thickness) and a mass spectrometer with a quadrupole mass filter and an electron multiplier detector. The GC-MS was operated in the electron ionization (EI) mode at 70 eV.

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Results

The results of the analysis of organochlorine compounds by GC-MS are shown in Figure 1. The figure shows the mass spectra of several organochlorine compounds, including DDT, DDE, and DDD. The mass spectra show the characteristic fragmentation patterns of these compounds.

Figure 1. Schematic diagram of the IBR setup.

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- Coin cells: dependent on Li^+ transport in carbonate electrolyte
 - weakly coordinating solvents
 - EMC
 - EC
- Breakdown can lead to H_2 production, flammability issues
- Diagnostic electrolyte analysis needed
 - Extremely limited sample volumes (a few μL)

BF MS analyses were conducted using a Benchtop Micro TOF-Orb, modified by the removal of the ESI source. A Hamilton 10 μL syringe equipped with a fused silica needle was used in the RF source. The needle was positioned approximately 0.5 cm from the aperture of the ion transfer capillary on the instrument, which was operated in the positive ion mode, using the standard "time-to-0" method.

RF is being used by the **US Army** for classified agent detection projects and **MRS R&D** w/ optional GoPro camera, J. Olyer, et al poster this meeting.

RF is being used for MS analysis of oligonucleotides. **NEU11** mRNA splicing w/ **UJ of Cincinnati** yields nanometer sensitivity for oligonucleotides

US Department of Energy is using RF in the field to analyze radio-active elements at **k levels**. — WITHOUT an KPI !

RF is being used to replace sampling using a MS from an OPERATING battery at a National Lab.

USE NIH, NIST & JEOL, have published that by using NiS for MALDI, SMS, LDH & DART that **MS sensitivity increases by 10-20 fold** (TITRALLY)

University of Wisconsin has used **RF** for single cell MALDI identifying its new nuclear proteins. We shoot cells into an ESJ at org./lab.

University of Illinois published that **RF can fly**! nanoparticles of liquids into levitated microparticles that yield less reaction kinetics.

For Abbott, **National LLC** used **RF** to detect PVA, w/ *av. MW* of 300,000, in pseudo 3D "printing" app

At Genentech, an n/wave demonstrates 20x improvement in MALDI sensitivity for proteins, peptides.

USE NIH is able to make electros. LC/MALDI for Seis; who offered to license LC ESMs for commercial client.

NIH, in it's first application for **RF**, PTM's of tubulin (Galland) were first tried, in actual brain cancer samples given a 100% sensitivity increase claim **NIH**

Seis offered to license **RF** for ESI/MS and for LC/MALDI 2006. Parallel 8-channel **RF** LC demoted with **SEIS**.

nano/morph morpho poly/polymer for Douglas and Spark-Holland's systems to parallel 384 or single channel microfluidic net detection, SPE, LC.

See more here: <http://www.national.com/nanotechnology212139ar.pdf> Some references: <http://nanotech.com/references/2014.pdf>

Examples: USE of HPLC, MS, APCI, MFC, VLE, UJ, USF, USF, USF, APC, AFPC, EDC and Hatch, Abbott, Biogen Idec, Genentech, Angen, Helix, Allergan, Spauld, Douglas, NIH, NIST, UCSD, InGa, Tech, Union, Douglas, NASA, Air Force, Navy, and Science offered to license. =

Just Shoot 'em in!

HPLC
UPLC
Syringe
Pipette
Chip
GS

GS

ESI

MS
MS/M
SIMS
Other

TIC a.

RNO₃(CMPD)₃⁺ b.

RNO₃(CMPD)₂⁺ c.

infusion time (sec)

Just Shoot 'em on!

nL = Excellent MALDI crystals.

Results & Discussion.

ESI = MASSIVE DILUTION ! Not programmable!

ESI

MS
MS/MS
IMS
Other

ESI < 1% in

IBF = 100 % (or less) into the ESI/MS !

IBF

"NO" DILUTION !

MS
MS/MS
IMS
Other

IBF 100% in, ...or less!

PROGRAMMABLE!

*Youngs-Golding, Ultrasmall-Lessons-Drops-Exhibit - Chavira, Jovanovski, Zhang, Li, Tapp, and Gonzalez-Schneider, Anal. Chem. 86(12), 4912-4916 (2014)
DOI: 10.1021/acs.analchem.5b00141
Copyright © 2014 American Chemical Society, <http://dx.doi.org/10.1021/acs.analchem.5b00141>

Figure 1 consists of two mass spectra, (A) and (B), showing relative abundance versus mass-to-charge ratio (m/z). Both spectra are for 20-mer oligonucleotides. Spectrum (A) is for the 20-mer TG (5'-ACA TCC CTC CGA CGG CTA TG-3') and spectrum (B) is for the 20-mer AG (5'-CGG CGG TGG TGC TTG-3'). Both spectra show a series of peaks corresponding to the oligonucleotide sequence. The x-axis ranges from 800 to 1200 m/z, and the y-axis represents relative abundance from 0 to 100. Insets in both spectra show a zoomed-in view of the region from m/z 1427 to 1527, highlighting peaks for A+K, A+L, and A+M.

Chemical structures of eight phosphazene compounds are shown, arranged in two rows of four. Each structure is a cage-like molecule with phosphorus (P) and nitrogen (N) atoms forming the core, and various organic groups attached to the phosphorus atoms.

- (5.1) MW 459: $\text{C}_2\text{H}_5\text{O}$ and OCH_2CF_3 groups.
- (5.2) MW 513: $\text{C}_2\text{H}_5\text{O}$ and OCH_2CF_3 groups.
- (5.3) MW 567: $\text{C}_2\text{H}_5\text{O}$ and OCH_2CF_3 groups.
- (5.4) MW 621: $\text{C}_2\text{H}_5\text{O}$ and OCH_2CF_3 groups.
- (5.5) MW 675: $\text{F}_3\text{CH}_2\text{CO}$ and OCH_2CF_3 groups.
- (5.6) MW 729: $\text{F}_3\text{CH}_2\text{CO}$ and OCH_2CF_3 groups.
- (5.7) MW 729: $\text{F}_3\text{CH}_2\text{CO}$ and OCH_2CF_3 groups.
- (5.8) MW 729: $\text{F}_3\text{CH}_2\text{CO}$ and OCH_2CF_3 groups.

In all experiments given here are an analog or programmable inductive energy source was employed to energize a fluid in a Gaussian surface. The Gaussian surfaces could be a capillary, a syringe, a pipette, a chip, an LC or UPLC columns or related device.

Typically a pneumatic flow was inductively energized in a programmed manner, to fly the liquid to a target in the nL/sec flow rate.

Circuitry was such that a drop or droplet flow was pushed or pulled to a target via appropriate connections, and optionally programmed WAVES.

Targets included MALDI plates, ESI sources or other targets from humans to inanimate objective to scientific instruments.

Fluids can be liquids of all types including whole human blood, polymers, lipids, peptides, proteins, oligonucleotides, etc.

Electric fields applied to said fluids *in a programmed manner* with appropriate circuitry to effect *rapid, directed* sample placement.

A general scheme is given below.

Note, droplets can be programmed to be pushed or pulled as connected and energized in appropriate waves.

IBF: Gaussian Surface.

$I \propto n^*$

* If linear

“Zero”

MS
MS/MS
IMS

Droplets shot into an ESI at ca. 1 m/s yield excellent SENSITIVITY because....

100% introduction efficiency.

Signal compression, (fast drops, more moles in less time) $dn/dt > \text{a diluting spray}$.)

High reproducibility, i.e., precise dn/dt , yields highly reproducibility d/dt . Like Printing.

Signal from a “zero” background.

ATTRIBUTES of IBF Droplets.

Programmable

Non-touch, directed dispense over a distance,

Highly Parallel Dispensing

Viscous liquid dispensing.

Multiple dispensing devices, gaussian surfaces, can be used.

Millisecond responses: nL, μ L and PL volumes w/ varied rate

Droplets appear to desolvate better than sprays increasing the sensitivity of even oligonucleotides [21].

[illegible]

More ethoxylated congeners (5,1), (4,2), and (3,3) lithium, and hydrate

- (4,2) and (3,3) lithium, form solvent complexes
- (2,4) and (1,5) protonate and lithium, form solvent complexes

Mass spectrum of the lithium salt of the 1,5-diol. The x-axis is m/z from 100 to 300, and the y-axis is relative intensity from 0.0 to 5.0×10^5 . The base peak is at m/z 250. Other significant peaks are at m/z 268, 286, 298, 274, 252, 234, 216, 198, 170, 152, 134, 116, 98, and 80. A blue arrow points to the peak at m/z 250, and a red arrow points to the peak at m/z 268.

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1B Theory

One 2D device, a flowing or droplet system. There are many EBF embeddings

The liquid volume is preserved, the flux is given by the Hagen-Poiseuille equation. The volume of fluid (V) that flows down a cylindrical pipe with radius a and length L is proportional to the radius after rate (η), the pressure pushing the fluid down the tube (P), the length of the tube (L), and the viscosity of the fluid. Here is how we do it:

$$V = (\pi r^2) \rho P \eta L$$

Since electric forces can be easily thought of and η with gravitational forces, and since $F = qE$, the forces on lipid drops can be changed rapidly. Because if F is a vector, we can deflect the drop as well.

$$F = qE$$

For a charged drop with initial velocity, u_0 , at a viscosity time, t , where $\eta = \text{size}/L$ where L is the selective contact of the region, η is the relative permeability and η is the solution conductivity and to this we have a question:

$$q = q_0 e^{(t/\eta)}$$

Now, a charged lipid drop on an electric field, cannot experience the force of E in x and z of course. Experimenters, different forces as well as the absorption η , η and η can be determined by the specific of the system. Drops are charged, and we can control them, we can control the forces electric, drag, buoyancy, gravity and controlling adding an drop to these adding the direction x , y , z .

$$F_x = m(a_x) = m(d^2x/dt^2) = F_{\text{electric}} + F_{\text{drag}} + F_{\text{buoyancy}} + F_{\text{gravity}} + F_{\text{col x}}$$

The same can be written for the y and z coordinates. Then, with possible model equations for F and η , we can actually calculate the trajectories of the entire system (electric, drag, buoyancy and gravity) and η . Answering the value and the direction of the forces.

$$V_1 = dv_1/dt, V_2 = dv_2/dt, V_3 = dv_3/dt$$

$$V_1^2 = V_1^2 + V_2^2 + V_3^2$$

$+f$ -Droplets follow the same trajectories, 17.

The graph shows the relationship between droplet size and time. The y-axis is 'Droplet Size (nm)' ranging from 0 to 150. The x-axis is 'Time (ms)' ranging from 0 to 1800. Experimental data points are shown as blue dots. A solid black line represents the 'Sample Theoretical Curve'. A dashed black line represents the 'Theoretical Curve with $\eta = 0.001$ '. A legend in the top right corner identifies the data series: 'Actual data points' (blue dots), 'Sample Theoretical Curve' (solid black line), and 'Theoretical Curve with $\eta = 0.001$ ' (dashed black line). The theoretical curves are defined by the equation $d = d_0 e^{-(t/\eta)}$. For the sample curve, $d_0 = 150$ nm and $\eta = 0.001$ ms. For the dashed curve, $d_0 = 150$ nm and $\eta = 0.0001$ ms.

The graph shows the relationship between droplet size and time for different viscosity values. The y-axis is 'Droplet Size (nm)' ranging from 0 to 150. The x-axis is 'Time (ms)' ranging from 0 to 1800. Experimental data points are shown as blue dots. Three theoretical curves are plotted: a solid black line for $\eta = 0.001$ ms, a dashed black line for $\eta = 0.0001$ ms, and a dotted black line for $\eta = 0.00001$ ms. A legend in the top right corner identifies the data series: 'Actual data points' (blue dots), 'Sample Theoretical Curve' (solid black line), 'Theoretical Curve with $\eta = 0.001$ ' (dashed black line), and 'Theoretical Curve with $\eta = 0.0001$ ' (dotted black line). The theoretical curves are defined by the equation $d = d_0 e^{-(t/\eta)}$. For the sample curve, $d_0 = 150$ nm and $\eta = 0.001$ ms. For the dashed curve, $d_0 = 150$ nm and $\eta = 0.0001$ ms. For the dotted curve, $d_0 = 150$ nm and $\eta = 0.00001$ ms.

Cell Phone pic of droplet MS Data, Note: 1. reproducible peak shape, 2. excellent peak symmetry with 3. rapid clear out time (ca. < 3 sec.) and with 4. low dead volume, here 5 sec/150 nL, can go 10-20x faster for protein therapeutic or other QC analysis.

ESI Data, $i = \text{signal} + \text{noise}$

IBF Data, $i = \text{signal} + \text{noise}$

150 nL

Total Ion current

100000

time

Bruker micro tech., w/ nanoLiter Cool Wave, droplet dispenser/3sec at flow ca. 70 nL/sec. Sample 10 Hz at ca. 150-1500 amu. TIC CMPO acquisition, 16Hz.

IBF = 100% into the ESI. PLUS, it does so from the lowest background @ the fastest dn/dt rate.

Droplets shot onto a non-conductor from the nL Programmable Wave.

1. Push-mode: the drop is same polarity as applied voltage, pull-mode: drop is opposite polarity, hence attraction.
2. Push-mode, if you touch plunger with skin (or apply any ground) NO dispersion will occur. the length of liquid column becomes conductive, presumably due to polarization.
3. Push-mode, I have used push mode to shoot drops straight up, no target except for ceiling 5ft away.
4. IBF Energy waves are useful.
5. Ongoing work just started. Demmed at Stanford elsewhere.

Droplet spread reduced by programming energy wave: 0.5; 1.0 & 5.0 sec.

Wave over 0.5 inch travel 1.0 sec 5.0 sec

Droplet dispersion is reduced by 5-10x over the 5 inch travel when energy is programmed. Best wave, timing, etc. TBD

- Temporal profiles
 - 100 nL, manually launched
 - 40 nL, field induced

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$$\begin{aligned}
 & \text{Note: equations for mass to charge ratio of a charged droplet equations, trajectory equations,} \\
 & \text{coulombic force equations, with geometric and velocity equations and drop geometric relationships.} \\
 & \begin{aligned}
 \frac{dQ}{dt} &= -\frac{2\pi\epsilon_0 r_p}{\rho_l} \frac{dV}{dt} \left(\frac{1}{r_p} + \frac{1}{r_c} \right) & \frac{dV}{dt} &= -\frac{4\pi}{3} \frac{d}{dt} \left(r_p^3 + r_c^3 \right) \frac{\rho_l}{2} \sum_{i=1}^N \frac{Q_i}{r_i} \cos \alpha_i \\
 & & & \frac{dV}{dt} &= -\frac{4\pi}{3} \frac{d}{dt} \left(r_p^3 + r_c^3 \right) \frac{\rho_l}{2} \sum_{i=1}^N \frac{Q_i}{r_i} \cos \alpha_i + \pi r_p^2 \\
 \cos \alpha_i &= \frac{r_c}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} & \cos \alpha_i &= \frac{r_c - r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} \\
 \sin \alpha_i &= \frac{r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} & \sin \alpha_i &= \frac{r_p - r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} \\
 \cos \alpha_i &= \frac{r_c - r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} & \cos \alpha_i &= \frac{r_c - r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} \\
 \sin \alpha_i &= \frac{r_p - r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}} & \sin \alpha_i &= \frac{r_p - r_p}{\sqrt{(r_c - r_p)^2 + (r_p - r_p)^2 + (r_c - r_p)^2}}
 \end{aligned} \\
 & \begin{aligned}
 r_i &= \sqrt{\left(\frac{r_p}{2} \right)^2 + \left(\frac{r_p}{2} \right)^2} & r_i &= \left(\frac{r_p}{2} + \frac{r_p}{2} \right) \\
 \frac{dQ}{dt} &= -\frac{2\pi\epsilon_0 r_p}{\rho_l} \frac{dV}{dt} \left(\frac{1}{r_p} + \frac{1}{r_c} \right) & \frac{dQ}{dt} &= -\frac{2\pi\epsilon_0 r_p}{\rho_l} \frac{dV}{dt} \left(\frac{1}{r_p} + \frac{1}{r_c} \right)
 \end{aligned}
 \end{aligned}$$

Drops travel like charged bullets Drops can be shot at ca. 1 m/s up to 1-2 m via the US Army APG-X1 Dispenser, 3 X 50 nLs, see target below. After "invention" in 2015, the US Army tested the dispensing accuracy of the Device by using GCMS.

See Jon Ayler ad, poster this meeting.

US ARMY OPG-ND, TWO MESSAGES FROM ABERDEEN PROVING GROUND,ND to nanoInk LLC. August 3, 2015 PARTIAL MESSAGE 1

Classification: UNCLASSIFIED, Caveats: NONE H Drew, Got your voice mail today. The nl pipette is working well. We have a procedure to fill it and dispense the volumes we need. We are preparing for a validation run using methyl salicylate. We plan to pipette volumes of 5 to 500 nL using 5 replicates. Will quantitate using GCMS. We will keep you posted on our progress.

Classification: UNCLASSIFIED, Caveats: NONE , Unidentified Project Leader August 24, 2015, PARTIAL MESSAGE 2 Classification:

UNCLASSIFIED, Caveats: NONE We have run curves and controls with very good results to a concentration range from 50 mg/ml to 15 mg/mL. We ran the controls at two concentrations with three replicates each. Accuracy and precision measures were very good (CVs <5%). We are repeating a stability study today tomorrow that we started on Friday just to ensure that we have good data going over a five day time period. It looks like we are almost there.

Unidentified Agent Accountable Officer



Can vary frequency, E, volume, wave form & placement.

Distance max = ca. 1.5 m.

Best drop accuracy, means one must direct the droplet, but align the droplets initial position.

Note if not aligned, droplets can wobble making the transfer to the MS, less precise than it might otherwise be.

WOBBLE

Gravity pulls droplet down.

Top: drop wiggles when shot ballistically.

Bot: timed E wave aligns drop first.

Programmed E reduces the WOBBLE

$E_0 = V = 0$

$E_1 = V f(t_1)$

$E_2 = V f(t_2)$

Applied field straightens up the drop, and launches it from same x, y, z position at "same" time done properly.

Different functions, rates available! Work ongoing. See below.

- Nanodroplet analysis enables glimpses at the intrinsic complex-formation chemistry of the phosphazene cyclotrimers
- Carbonate complexes increase with increasing numbers of $-\text{OCH}_2\text{CF}_3$ moieties
- Hydrates are preferred for compounds with more $-\text{OC}_2\text{H}_5$ groups
- Useful approach for systems with very limited sample volume

"ANY" Gaussian Surface + ELECTRIC INDUCTION
can be an ION SOURCE or aMALDI), SIMS, LDI or other DISPENSER, that's optionally Android controlled.

Finally, as given at ASMS's Asilomar Meeting in 2016 with the University of Cincinnati and Idaho National Laboratory, 100% input efficient (BF based) UPLC MS of nucleosides yielding similar sensitivities has been observed on the same platform that executes 100% input efficient, millisecond infusion sample input.

Work continues.

24. Robert L. Ross, M. Jora, Andrew D. Sauter Jr., Andrew D. Sauter III and Patrick A. Limbach, Droplet Based Sampling of RNA Hydrolasates by Induction Based Fluidics, presented at the October 2016 ASMS Asilomar Meeting, Monterey, CA.

25. Robert L. Ross, M. Jora, Patrick A. Limbach, G. S. Groenewold, Andrew D. Sauter III and Andrew D. Sauter Jr., A Single Programmable Android Controlled Energy Embodiment for MALDI, SIMS, LDI and ESI, presented at the October 2016 ASMS Asilomar Meeting, Monterey, CA.